

High-throughput colorimetric assays for mercury(II) in blood and wastewater based on the mercury-stimulated catalytic activity of small silver nanoparticles in a temperature-switchable gelatin matrix†

Cite this: *Chem. Commun.*, 2014, 50, 9196

Received 20th May 2014,
Accepted 25th June 2014

DOI: 10.1039/c4cc03851g

www.rsc.org/chemcomm

Zongzhao Sun, Ning Zhang, Yanmei Si, Shuai Li, Jiangwei Wen, Xiangbing Zhu and Hua Wang*

A catalysis-based, label-free, and high-throughput colorimetric protocol has been initially proposed for detecting mercury(II) in blood and wastewater with 96-cell plates, based on the mercury-enhanced catalytic activity of small silver nanoparticles synthesized in a gelatin matrix with unique temperature switchable sol–gel transition.

Recent years have witnessed the rapid development of nano-scale materials of noble metals (*i.e.*, Au, Ag, Pt, and Pd) with many fantastic physicochemical properties.¹ Especially, their unique enzyme-like catalysis has attracted considerable interest in catalysis and biochemical analysis applications. For example, gold nanoparticles (NPs) could present peroxidase-like catalysis activity for sensing H₂O₂ and glucose.² Catalytic Pt NPs have also been utilized for the detection of H₂O₂ and scavenging superoxide free radicals.² Moreover, the catalytic activity of noble metal nanomaterials can substantially depend on their sizes, known as the “size effect”, for example, high catalytic activity can be observed for gold NPs with sizes smaller than 5.0 nm.³ As a result, many efforts have been devoted to the synthesis of size-reducing gold NPs or nanoclusters.⁴ The preparation of cheap and catalytic small silver NPs (AgNPs), however, still remains a challenging but abstractive issue, due to the difficult synthesis, low stability, and the poor catalytic performance of small AgNPs.

As the most hazardous heavy metal ions, mercury ions were targeted by many modern detection methods such as DNA folding-based electrochemical detection, fluorescence quenching-based analysis, and gold or silver NPs-based colorimetric assay.⁵ However, these methods might suffer from either low detection sensitivity and throughput, or poor analysis stability and abilities against background interferences. Recently, some catalytic noble metal nanomaterials have been utilized for probing heavy metal

ions like Hg²⁺ ions that could inhibit and stimulate their catalytic activity through a specific Au⁺–Hg²⁺ interaction and the change in the surface properties of AuNPs or AgNPs by forming Ag–Hg alloys, respectively.⁶ Such a catalysis-based analysis methodology may possess some advantages over the traditional methods like fluorescence-based ones in terms of the detection stability and targeting abilities against interferences from the complicated media (*i.e.*, blood).

Gelatin, as a natural biopolymer derived from collagen widely used in food products and medicines,⁷ is nontoxic, non-immunogenic, biodegradable, and especially with unique sol–gel transition (critical transition at 35 °C). Such a biomaterial was introduced for the synthesis of AgNPs but entailing additional reductants like maltose and citrate.⁸ Also, small AgNPs with Hg²⁺-enhanced catalysis activity are rare. In this work, the gelatin matrix was employed to act as the reducing agent and the stabilizing matrix for the one-pot synthesis of small AgNPs at physiological temperature (37 °C) without adding any reducing agents. Unexpectedly, the prepared gel-AgNPs could display Hg²⁺-stimulated powerful catalytic activities in catalyzing 3,3',5,5'-tetramethylbenzidine (TMB)–H₂O₂ reactions.

The special phenomenon of mercury(II)-enhanced catalysis of small gel-AgNPs was explored using comparable tests in the absence and presence of Hg²⁺ ions (Fig. 1A). It was found that gel-AgNPs showed a lower catalytic activity in the TMB–H₂O₂ reaction at a meaningfully low concentration (Fig. 1A(c)). However, their catalytic activity could be significantly enhanced by Hg²⁺ ions, showing a deep blue reaction product in the photograph (Fig. 1A(d)). Furthermore, effects of some other ions on stimulating the catalytic activity of gel-AgNPs were investigated (Fig. 1B and Fig. S1, ESI†). One can observe that common inorganic ions do not have the ability to stimulate the catalytic activity of gel-AgNPs, even at concentrations of about 100-fold higher than that of Hg²⁺ ions. Based on the specific mercury-stimulated catalytic activity of gel-AgNPs, a label-free colorimetric protocol has been thus proposed for the detection of Hg²⁺ ions. Scheme 1 illustrates the catalysis-enhanced analysis

Shandong Province Key Laboratory of Life-Organic Analysis, School of Chemistry and Chemical Engineering, Qufu Normal University, Qufu City, Shandong Province 273165, P. R. China. E-mail: huawangqfnu@126.com; Fax: +86 5374456306; Tel: +86 5374456306

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c4cc03851g

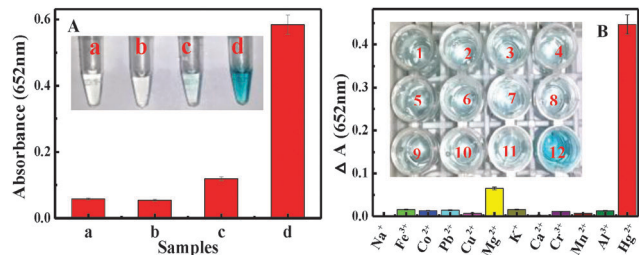


Fig. 1 (A) Comparison of catalysis activity in TMB–H₂O₂ reactions among (a) water (control), (b) 8.0×10^{-7} M Hg²⁺, (c) 7.2×10^{-6} M gel-AgNPs, and (d) 8.0×10^{-7} M Hg²⁺ with 7.2×10^{-6} M gel-AgNPs, corresponding to colorimetric photographs (inset); (B) investigation of the catalytic activity of gel-AgNPs in TMB–H₂O₂ reactions stimulated by various metal ions Na⁺, Fe³⁺, Co²⁺, Pb²⁺, Cu²⁺, Mg²⁺, K⁺, Ca²⁺, Cr³⁺, Mn²⁺, Al³⁺ (each 5.0×10^{-5} M), and Hg²⁺ (4.5×10^{-7} M), with corresponding colorimetric photographs (inset, samples 1–12).

protocol using 96-well plates towards high-throughput colorimetric assays for Hg²⁺ ions in water, blood, and wastewater. As described in Scheme 1, gel-AgNPs (thawed at 37 °C) alone (left top) exhibited no significant catalysis in TMB–H₂O₂ reactions at low concentration. In contrast, the catalysis activity of gel-AgNPs could be greatly enhanced by Hg²⁺ ions to yield blue reaction products.

A transmission electron microscope (TEM) and a UV-vis spectrophotometer were separately utilized to characterize gel-AgNPs in the absence and presence of mercury(II) (Fig. 2). As shown in Fig. 2A, original gel-AgNPs were dense and well dispersed with the average particle diameter of about 5.0 nm (Fig. 2A, left). When Hg²⁺ ions were introduced, AgNPs in the suspension became sparser and smaller (Fig. 2A, right), as more apparently shown in the amplified view of one particle (inset, right top). Also, the hydrodynamic diameters of gel-AgNPs before and after Hg²⁺ treatment were comparably examined by dynamic light scattering (Fig. S2, ESI[†]), showing that their average sizes decreased from about 13.0 nm to 8.5 nm. Furthermore, UV-vis spectra were also recorded for gel-AgNPs with and without Hg²⁺ ions (Fig. 2B(a)). It was observed that Hg²⁺ ions could increase

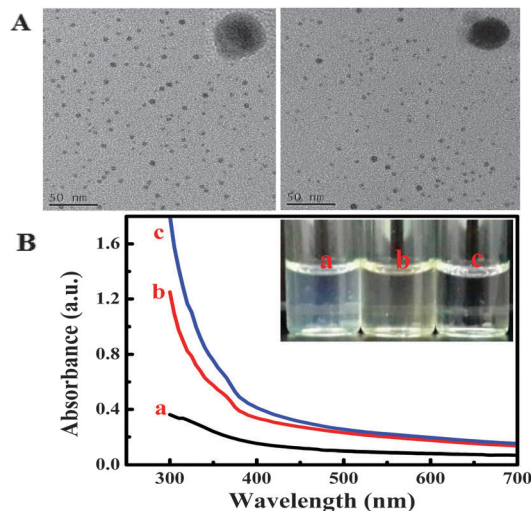
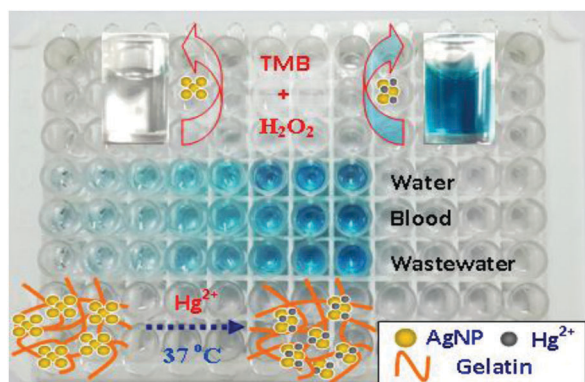


Fig. 2 Characterization of (A) TEM images of gel-AgNPs alone (left) and gel-AgNPs with Hg²⁺ (right), each with an amplified view of one particle (inset), and (B) UV-vis spectra of (a) gel, (b) gel-AgNPs alone, and (c) gel-AgNPs with Hg²⁺, corresponding to the photographs (inset).

the UV-vis absorbance of gel-AgNPs; yet, they could cause the yellowish gel-AgNPs to fade out, as evidenced from their photographs (inset, Fig. 2B(b) and (c)). The above results indicate that the mercury(II) etching of AgNPs in gel-AgNPs could occur when they were treated with Hg²⁺ ions.

Furthermore, the enhanced catalysis mechanism for the Hg²⁺-stimulated catalytic activity of gel-AgNPs was verified by essential experiments to make sure whether mercury(II) could etch gel-AgNPs. Herein, gel-AgNPs were deliberately treated under UV lamp so as to grow AgNPs till their yellowish color changed to deep yellow. Fig. S3A (ESI[†]) shows TEM images of the UV-treated gel-AgNPs before and after the Hg²⁺ addition. A size-reducing change of AgNPs was observed for the latter. It could also be verified by the UV-vis spectra showing that UV-treated gel-AgNPs could decrease in size after Hg²⁺ etching, as shown in the corresponding photographs (inset) (Fig. S3B, ESI[†]). On the basis of the above evidence, the mercury(II)-stimulated enhancement of the peroxidase-like catalytic activity of gel-AgNPs should result mainly from the decreased size of AgNPs by Hg²⁺ etching, of which the size-reducing AgNPs might achieve much higher catalysis activity, known as the “size effect”, as also evidenced elsewhere for gold and Fe₃O₄ nano-materials.³ Also, the special interaction between Hg²⁺ and AgNPs and the formation of Ag–Hg alloys changing the surface properties of AgNPs might be involved,⁶ which remains to be further investigated in the future.

Moreover, the mercury(II)-enhanced catalytic dynamics of gel-AgNPs was investigated in the absence and presence of mercury(II) (Fig. S4, ESI[†]). Here, colorimetric measurements were performed alternatively for different concentrations of TMB and H₂O₂ to obtain the Michaelis–Menten curves⁹ (Fig. S4B and D, ESI[†]). It was found that Hg²⁺-stimulated gel-AgNPs presented a much lower Michaelis constant (K_m) than gel-AgNPs alone for TMB and H₂O₂, together with larger maximal



Scheme 1 Colorimetric assays of Hg²⁺ in different samples by the TMB–H₂O₂ reactions catalyzed by gel-AgNPs with Hg²⁺-enhanced catalytic activity, where different concentrations of Hg²⁺ ions in water, blood, and wastewater were measured using 96-well plates with gel-AgNPs.

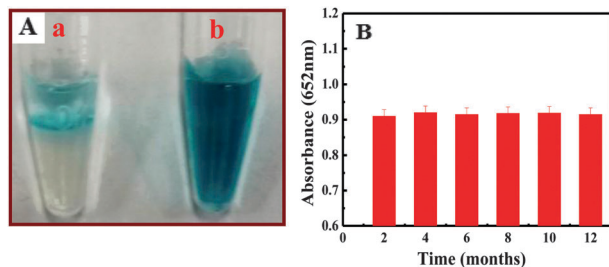


Fig. 3 (A) Comparable photographs of the catalytic activity of gel-AgNPs in TMB–H₂O₂ reactions with gel-AgNPs originally (a) stored at 4 °C, and (b) thawed at 37 °C with Hg²⁺ ions; (B) mercury(II)-stimulated catalytic stability of gel-AgNPs stored at 4 °C in a fridge over different time periods.

reaction velocity (V_{\max}). The data indicate that Hg²⁺-stimulated gel-AgNPs could possess stronger catalytic activity especially higher affinity to TMB and H₂O₂. Colorimetric analysis of Hg²⁺ ions could thereby be achieved based on the Hg²⁺-enhanced catalysis of gel-AgNPs.

Moreover, the unique property of sol-gel transition of the gelatin matrix (critical temperature at 35 °C) was investigated in switching the catalysis of gel-AgNPs (Fig. 3). It was noted that gel-AgNPs in the gel phase (at 4 °C) with Hg²⁺ ions could not catalyze the TMB–H₂O₂ reactions, where the catalytic reactions occurred only on the gel surface (Fig. 3A(a)). When gel-AgNPs were thawed at 37 °C and diluted to be further mixed with Hg²⁺ ions, they could exert strong catalysis for the colorimetric reaction (Fig. 3A(b)), with the Hg²⁺-stimulation time of about 5.0 min (Fig. S5, ESI[†]). Particularly, such a temperature-switching catalysis of gel-AgNPs offered by the gelatin matrix could allow for high catalysis stability of gel-AgNPs, remaining up to one year without a significant change (Fig. 3B). Importantly, gel-AgNPs could be stably immobilized onto 96-well plates for long-term storage at 4 °C to facilitate colorimetric Hg²⁺ assays in a label-free and high-throughput way. An advantage over other mercury(II) sensing assays, including those based on mercury-stimulated catalysis activity reported elsewhere,⁶ could thus be expected.

Moreover, the dosage of gelatin for the synthesis of gel-AgNPs was optimized as 4.0% (Fig. S6A, ESI[†]). Furthermore, the reaction conditions for the colorimetric Hg²⁺ analysis were optimized as 7.2 μM gel-AgNPs (Fig. S6B, ESI[†]), ionic strengths of 5.0 mM KNO₃, a color reaction time of 20 min, neutral solution (pH 6.0–8.0) at room temperature (10–25 °C) (Fig. S7, ESI[†]), with 0.25 mM TMB and 10 mM H₂O₂ (Fig. S4A and C, ESI[†]).

Finally, calibration detection curves were obtained for Hg²⁺ ions with different concentrations in water, blood, and wastewater samples using 96-well plates (Fig. 4), corresponding to photographic results shown in Scheme 1. It was found that the typical UV-vis absorbance values could increase with increasing Hg²⁺ concentrations (Fig. 4A). A linear detection range of Hg²⁺ concentrations was obtained from 0.50 to 800 nM with the limit of detection (LOD) being 0.125 nM in water (Fig. 4B), which is lower than those of most other detection methods (Table S1, ESI[†]). Moreover, the colorimetric detection of Hg²⁺ ions in blood and wastewater was conducted, with linear Hg²⁺ concentrations ranging from 5.0 nM to 700 nM and 1.0 nM to 800 nM, respectively. High detection selectivity for Hg²⁺

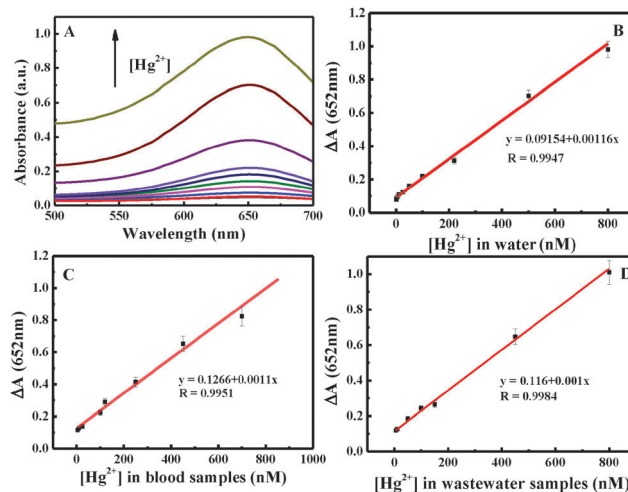


Fig. 4 Calibration detection curves for the linear relationship of absorbance values vs. different concentrations of Hg²⁺ ions in samples of (B) water, (C) blood, and (D) wastewater, with typical UV-vis spectra of reaction products shown in (A). Colorimetric assays of Hg²⁺ ions in different samples were conducted using TMB–H₂O₂ reactions, corresponding to photographic results shown in Scheme 1.

ions could also be achieved with no significant interference from other metal ions, as shown in Fig. 1A. Therefore, the developed catalysis-based colorimetric assays could allow for the high-throughput Hg²⁺ detection with pretty high sensitivity and selectivity.

In summary, a temperature-switchable gelatin matrix has been successfully employed for the one-pot synthesis of catalytic small AgNPs without any reductants. The resulting gel-AgNPs could display mercury-stimulated catalysis activity toward a simple, rapid, label-free, and high-throughput colorimetric protocol for probing mercury(II) in blood and wastewater using 96-cell plates. The catalysis-based detection mechanism involved was thought to result mainly from the mercury etching of AgNPs in the gelatin matrix leading to smaller sizes of AgNPs with greatly enhanced catalysis activity for TMB–H₂O₂ reactions. The unique property of sol-gel transition of the gelatin matrix could not only facilitate the temperature-switchable catalysis of gel-AgNPs, but also allow for high catalysis stability of gel-AgNPs. Remarkably, the catalysis-based method could detect Hg²⁺ ions in complicated wastewater and blood with high sensitivity, selectivity, and throughput. It might also circumvent some disadvantages of traditional detection approaches like fluorescence assays in terms of the detection stability and abilities against interferences from the complicated media. Such a colorimetric mercury assay promises huge potential applications in the clinical diagnosis, environmental monitoring, and pharmaceutical analysis fields.

This work is supported by the National Natural Science Foundations of China (No. 21375075, 21302109, and 21302110) and the Taishan Scholar Foundation of Shandong Province, China.

Notes and references

- (a) H. You, S. Yang, B. Ding and H. Yang, *Chem. Soc. Rev.*, 2013, **42**, 2880; (b) Ö. Metin, S. F. Ho, C. Alp, H. Can, M. N. Mankin, M. S. Gültekin, M. Chi and S. Sun, *Nano Res.*, 2013, **6**, 10.

- 2 (a) Y. Jv, B. Li and R. Cao, *Chem. Commun.*, 2010, **46**, 8017; (b) T. Hamasaki, T. Kashiwagi, T. Imada, N. Nakamichi, S. Aramaki, K. Toh, S. Morisawa, H. Shimakoshi, Y. Hisaeda and S. Shirahata, *Langmuir*, 2008, **24**, 7354; (c) M. Ma, Y. Zhang and N. Gu, *Colloids Surf., A*, 2011, **373**, 6.
- 3 (a) F. F. Peng, Y. Zhang and N. Gu, *Chin. Chem. Lett.*, 2008, **19**, 730; (b) S. Lee, C. Fan, T. Wu and S. L. Anderson, *J. Am. Chem. Soc.*, 2004, **126**, 5682; (c) D. Matthey, J. Wang, S. Wendt, J. Matthiesen, R. Schaub, E. Lægsgaard, B. Hammer and F. Besenbacher, *Science*, 2007, **315**, 1692; (d) G. R. Bamwenda, S. Tsubota, T. Nakamura and M. Haruta, *Catal. Lett.*, 1997, **44**, 83.
- 4 X.-X. Wang, Q. Wu, Z. Shan and Q.-M. Huang, *Biosens. Bioelectron.*, 2011, **26**, 3614.
- 5 (a) L. Yang, C. Zhang, H. Jiang, G. Li, J. Wang and E. Wang, *Anal. Chem.*, 2014, **86**, 4657; (b) J. Xie, Y. Zheng and J. Y. Ying, *Chem. Commun.*, 2010, **46**, 961; (c) M. Annadhasan, T. Muthukumarasamyvel, V. Sankar Babu and N. Rajendiran, *ACS Sustainable Chem. Eng.*, 2014, **2**, 887.
- 6 (a) R. Zhu, Y. Zhou, X.-L. Wang, L.-P. Liang, Y.-J. Long, Q.-L. Wang, H.-J. Zhang, X.-X. Huang and H.-Z. Zheng, *Talanta*, 2013, **117**, 127; (b) Y.-J. Long, Y.-F. Li, J.-J. Zheng and C.-Z. Huang, *Chem. Commun.*, 2011, **47**, 11939; (c) G.-L. Wang, X.-F. Xu, L.-H. Cao, C.-H. He, Z.-J. Li and C. Zhang, *RSC Adv.*, 2014, **4**, 5867.
- 7 (a) P. H. von Hippel and K.-Y. Wong, *Science*, 1964, **145**, 577; (b) A. Veis, *Int. Rev. Connect. Tissue Res.*, 1964, **3**, 113.
- 8 (a) O. S. Oluwafemi, Y. Lucwaba, A. Gura, M. Masabeya, V. Ncapayi, O. O. Olujimi and S. P. Songca, *Colloids Surf., B*, 2013, **102**, 718; (b) K. Mahmoud and M. Abbo, *Spectrochim. Acta, Part A*, 2013, **116**, 610.
- 9 L. Gao, J. Zhuang, L. Nie, J. Zhang, Y. Zhang, N. Gu, T. Wang, J. Feng, D. Yang and S. Perrett, *Nat. Nanotechnol.*, 2007, **2**, 577.